

**1219-Pos****Discrete and Continuous Three Dimensional Simulations for Fluorescence Recovery in Bacteria**

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Recent evidence indicates that components of functional molecular complexes in living cells may turnover relatively rapidly over timescales of seconds. Experimental methods such as fluorescence recovery after photobleaching (FRAP) may be used to probe the kinetics of the dynamic turnover process. Several models exist to model the FRAP process within a cell, however the majority of these require assumptions specific to the size and topology of eukaryotic cells. In the present work we present a robust discrete stochastic simulation of the FRAP process in three dimensional space for application to turnover processes in the bacterial cytoplasm. This is compared with a complementary continuous numerical simulation based on the finite element method (FEM). The effect of the dimensions and shape of the bacterial cell are analysed to elucidate the rôle of boundary effects, and low copy number régimes are simulated to study the effect of intrinsic noise in such systems. We show how these simulations may be used to optimise kinetic parameters based on experimental data.

**1220-Pos****pH Induced Dynamics Enables the Peptide Exchange of MHCII Molecules**Haruo Kozono<sup>1</sup>, Naoki Ogawa<sup>2</sup>, Osami Kanagawa<sup>2</sup>, Yuji Sasaki<sup>3</sup>.

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MHC II acquires antigenic peptide at acidic endosomes, which contain proteases and DM for peptide generation and peptide exchange reactions. The mechanism of peptide exchange reaction catalyzed by DM is still an enigma. We noticed that peptide exchange can be completed by just lowering pH for most MHC II isotypes. Thermal denaturation studies by differential scanning calorimeter have shown that the MHC II are more stable at pH5 than pH7, and the effect was entropy driven which imply that molecular flexibility at low pH has some role for the reaction. Thus, we carried out the diffracted X-ray tracking (DXT) analysis of peptide/MHC II, which should detect even a slight movement of the peptide/MHC II complex at the single molecule level. The molecular movement of peptide/MHC II complexes appeared to be larger at pH5 than pH7 when low affinity peptide was bound. This sustains results of the thermodynamic studies. In order to test the effect of the increased flexibility, we artificially restrained the movement of MHC molecule. When MHC II was produced as leucine-zipper rigidified form at the C-terminus, the movement was heavily restricted on DXT measurement. Furthermore, the rigidified MHC II molecule show reduced peptide exchange capability. These results entail that the flexibility at lower pH has a role for the peptide exchange reaction of MHC II molecules.

**1221-Pos****Molecular Mechanisms for Phosphorylation Driven Dissociation of Rb-E2F Complexes**Jason R. Burke<sup>1</sup>, Jeffrey G. Pelton<sup>2</sup>, Seth M. Rubin<sup>1</sup>.

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The Retinoblastoma Protein (Rb) functions as a negative regulator of cell growth in part by physically sequestering and repressing the transactivation activity of E2F. It is well established that phosphorylation of Rb by Cyclin Dependent Kinases disrupts binding between Rb and E2F, however it is unknown which of the 15 CDK consensus phosphorylation sites on Rb are required to disrupt the interaction between the pocket domain of Rb and the transactivation domain of E2F (E2FTD). In this work, we use calorimetric assays to reveal that phosphorylation at S608/S612 and T356/T373 are together sufficient to reduce the affinity of E2FTD for Rb pocket 250-fold, the same as fully-phosphorylated Rb. Nuclear Magnetic Resonance is used to identify the phosphorylation-dependent conformational changes that directly inhibit E2FTD binding. Specifically, we have found that phosphorylation of S608/S612 promotes intramolecular binding between the flexible pocket linker and the pocket domain of Rb, while phosphorylation at T356/T373 enhances binding between the N-terminal domain and pocket domain of Rb. Taken together, our results reveal two novel mechanisms for how phosphorylation of Rb modulates binding between E2FTD and Rb pocket, and we describe for the first time a function for the N-terminal domain in the inactivation of Rb.

**1222-Pos****Expanding the Range of Redox Potentials of the 2Fe-2S Clusters of the Outer Mitochondrial Membrane Protein MitoNEET**John A. Zuris<sup>1</sup>, Mark L. Paddock<sup>2</sup>, Andrea R. Conlan<sup>1</sup>, Edward C. Abresch<sup>2</sup>, Rachel Nechushtai<sup>3</sup>, Patricia A. Jennings<sup>1</sup>.

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MitoNEET is a recently discovered outer mitochondrial membrane protein that harbors a 2Fe-2S cluster bound to a unique 3Cys-1His coordination (1). We measured a pH-dependent redox potential with a value near 0 mV at pH 7.0 ( $E_{m,7}$ ). This value lies intermediate between most low potential 4Cys-coordinated ferredoxin-like (Fd) centers (~-300 mV) and most high potential 2Cys-2His-coordinated Rieske centers (~+300 mV) (2). Upon replacing the single His87 ligand with Cys, we obtained an  $E_{m,7}$  near -300 mV closer to that of Fd clusters (Figure). Upon replacing Lys55 located near His87 with Met,  $E_{m,7}$  increases to near +250 mV, closer to that of high potential Rieske clusters (Figure). This shows that there is a large interaction between Lys55 and His87 and that mitoNEET is robust to large changes in the  $E_{m,7}$  of its 2Fe-2S clusters. Thus, we have engineered stable mutant mitoNEET with  $E_{m,7}$  values over a range of almost 600 mV.

(1) Paddock et al. (2007) Proc Natl.

Acad. Sci USA 104, 14342-14347

(2) Meyer (2008) J Biol Inorg Chem 13, 157-170

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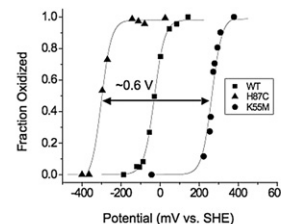


Figure. Spectroelectrochemical titrations of WT MitoNEET (squares) with H87C (triangles) and K55M (circles) mutants. The fraction oxidized versus solution potential was fit to a Nernst equation to obtain the redox potential. Measurements were done in 50 mM Tris 100 mM NaCl pH 8.0. All potential values were corrected to Standard Hydrogen Electrode (SHE).

**1223-Pos****"A Comparative Thermal Kinetic Investigation of the Fresh and Stale-Expired Protein Denaturing Transition"**

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Protein, in eggs, has a biological quality greater than any other natural food. The quality of the protein in eggs means that all egg protein can be used for synthesis and limits the amount burned as fuel or stored as fat. Hence, the quality of egg protein can be an important factor in weight loss and essential to understand more facts about egg proteins. In this study, a comparative thermal kinetic investigation of the fresh and stale-expired proteins was made using calorimetric technique. Three different scans as temperature scans, heating rate scans and time scans were performed using differential scanning calorimetry varying temperature range from 0 °C to 200 °C and heating ramp rates from 1 °C/min to 20 °C/min. Two protein samples were used; one obtained from a fresh egg and other from the stale-expired egg from the same batch. The denaturing transition for the stale egg protein occurred at the higher temperature with smaller enthalpy and needed a larger activation than the fresh protein. This indicates that stale-expired egg provides less energy and need more energy to be activated or burnt than the fresh egg protein. Hence, the stale protein would increase more fat and need more energy to burn if eaten in the food.

**Keywords:** Calorimetry, fresh and stale Protein, Kinetics, Activation Energy, Denaturing transition, thermodynamics

**1224-Pos****High-Order Correlations in Internal Protein Motions and Energetics**Arvind Ramanathan<sup>1</sup>, Andrej Savol<sup>2</sup>, Chris Langmead<sup>1</sup>, Pratul Agarwal<sup>3</sup>, S. Chakra Chennubhotla<sup>2</sup>.

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**Background:** Despite originating as a linear sequence of amino acids, folded proteins display a remarkable diversity and specificity of motions, or dynamics, which constitute the building blocks of cellular metabolism. A growing body of evidence also suggests that these motions are hierarchical, involving a multitude of spatial and temporal scales. A key task in biology is thus to elucidate the relationship between hierarchical nature of protein dynamics and function. Characterizing these spatial/ temporal fluctuations in molecular dynamics (MD) simulations is critical to understanding enzyme catalysis, ligand binding, and allosteric signaling - all therapeutically exploitable processes.

**Results:** Using 0.5us simulation for ubiquitin, a 76 residue protein that labels other proteins for degradation, we show that positional deviations exhibit non-Gaussian behavior (kurtosis >> 3), at functionally important regions of the protein. To analyze the spatial deviations we propose a general and statistically rigorous method Quasi Anharmonic Analysis (QAA) that meaningfully captures non-Gaussian behaviors overlooked with established methods. QAA,